

Supporting Information

Use of Selenium Bound Resin for the Solid Phase Synthesis of Substituted Isoxazolyl-Substituted (*E*)-Olefins

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1. Experimental section

The melting points were uncorrected. ^1H NMR (400 MHz) and ^{13}C NMR (100 MHz) spectra were recorded on a Bruker Avance 400 spectrometer in CDCl_3 with TMS as the internal standard; chemical shifts were quoted in ppm and *J* values were given in Hz. IR spectra were recorded on a Bruker Vector 22 spectrometer. EIMS were run on a HP 5989B mass spectrometer. Elemental analysis was run on Thermofinnigan Flash EA 1112. HPLC were run on a Shimadzu LC-6A. X-ray was run on a Rigaku RAXIS-RAPID diffractometer.

Chromatographic conditions(HPLC)

Column: ODS 5μ 250X4 mm. Mobile phase: THF/MeOH/ H_2O =51/17/32 (V/V).

Flow rate: 0.8mL/min. Detector: UV287nm.

Typical procedure for the preparation of isoxazole supported selenium resin 3:

To a suspension of the swollen polystyrene-supported selenenyl bromide (Br: 0.99mmol/g) resin **1** (2.5 g) in dry THF /DMF (V/V=5:1) (30 mL) was added NaBH₄ (5 mmol) under nitrogen atmosphere at 40°C. After stirring for 8 h at 40°C, propargyl bromide (5.5 mmol) was added dropwise under nitrogen atmosphere, and stirring for another 12 h. The resin **2** was collected by filtration, washed with THF (20 mL×2), MeOH (20 mL×2) and CH₂Cl₂ (20 mL×2) and dried in vacuum. To a suspension of the swollen resin **2** (2.5g) in CH₂Cl₂ was added a mixture of hydroximoyl halide (7.5 mmol) in 10 mL CH₂Cl₂ (prepared from 7.5 mmol of aldoxime and 7.5 mmol of NCS stirring at r.t. for about 3 hours when use). A mixture of Et₃N (15 mmol) in 15 mL CH₂Cl₂ was slowly dropwised in 3 portions every 8 hours (each time 5 mmol in 5 mL dry CH₂Cl₂ was added). After stirring for 24 h at r.t., The resin **3** was collected by filtration, washed with THF (20 mL×2), ether (20 mL×2), THF/H₂O (3:1) (20 mL×2), H₂O (20 mL×2), THF (20 mL×2), benzene (20 mL×2), MeOH (20 mL×2), and CH₂Cl₂ (20 mL×2), and dried in vacuum.

Typical procedure for the preparation of 3-aryl-5-*E*-substituted-ethenyl isoxazoles (products 5a-5g):

To a suspension of the swollen resin **3** (0.5 g) in dry THF, cooled to –60°C, was added dropwise LDA (2M in THF/hexane, 0.3 mL) under nitrogen. After stirring for 1.5 h at –60°C, a solution of allyl bromide (2 mmol) in 1 mL of dry THF was added. The suspension was stirred for another 0.5 h at –60°C. Slowly warm up to –40°C in 0.5 h then quenched with 1 mL H₂O. The resin **4** was collected by filtration and washed with THF (10 mL×2), THF/H₂O (3:1) (10 mL×2), H₂O (10 mL×2), THF (10 mL×2), and CH₂Cl₂ (10 mL×2). The washed resin was suspended in THF (15 mL), to the mixture was added 30% (aq.) H₂O₂ (0.5 mL) and stirred for 1 h at 0°C followed by 20 minutes at room temperature. The mixture was filtered and the resin was washed with CH₂Cl₂ (15 mL×2). The filtrate was washed with H₂O (30 mL×2), dried over MgSO₄, and evaporated to dryness in vacuum.

Typical procedure for the preparation of Isoxazolyl and Isoxazolinyl substituted (*E*)-olefins (products 8a-8k):

To a suspension of the swollen resin **6** (0.5 g) in CH₂Cl₂ was added a mixture of hydroximoyl halide (1.5 mmol) in 10 mL CH₂Cl₂ (prepared from 1.5 mmol of aldoxime

and 1.5 mmol of NCS stirring at r.t. for about 3 hours when use). A mixture of Et₃N (3 mmol) in 15 mL CH₂Cl₂ was slowly dropwised in 3 portions every 8 hours (each time 1 mmol in 5 mL dry CH₂Cl₂ was added). After stirring for 24 h at r.t., The resin **7** was collected by filtration, washed with THF (10 mL×2), ether (10 mL×2), THF/H₂O (3:1) (10 mL×2), H₂O (10 mL×2), THF (10 mL×2), benzene (10 mL×2), MeOH (10 mL×2), and CH₂Cl₂ (10 mL×2), and dried in vacuum. The washed resin was suspended in THF (15 mL), to the mixture was added 30% (aq.) H₂O₂ (0.5 mL) and stirred for 1 h at 0°C followed by 20 minutes at room temperature. The mixture was filtered and the resin was washed with CH₂Cl₂ (15 mL×2). The filtrate was washed with H₂O (30 mL×2), dried over MgSO₄, and evaporated to dryness in vacuum.

2. Characterization data of key compounds.

Purity and yield are determined by the crude product and NMR MS, FTIR and EA are determined by the purified product.

¹H NMR (400 MHz), ¹³C NMR (100 M Hz), MS, FTIR and EA of the products **5b**, **5c**, **5f**, **8c**, **8d**, **8j**, **8k**.

5b: 82.3mg, yellow low pointing solid; ¹H NMR (CDCl₃) δ 7.72 (2H, d, *J* = 8.2 Hz), 7.28 (2H, d, *J* = 8.2 Hz), 7.02 (1H, dd, *J*₁ = 10.4 Hz, *J*₂ = 15.6 Hz), 6.53-6.47 (3H, m), 5.53 (1H, d, *J* = 16.8 Hz), 5.17 (1H, d, *J* = 10.4 Hz), 2.43 (3H, s); ¹³C NMR (CDCl₃) δ 168.9, 163.0, 140.5, 136.2, 135.8, 130.0, 127.1, 126.6, 122.4, 117.4, 99.9, 21.8; MS *m/z* 158(100), 211(M⁺); IR ν_{max} (cm⁻¹) 3405, 3027, 1624, 1493, 1449, 967, 753, 692. Elemental analysis Calcd. for C₁₄H₁₃NO, C 79.59 %; H 6.20 %; N 6.63% Found C 79.38 %; H 6.29 %; N 6.72%.

5c: 81.4mg, pale yellow solid, mp. 143-145°C; ¹H NMR (CDCl₃) δ 7.72 (2H, d, *J* = 7.6 Hz), 7.55 (1H, d, *J* = 16.0 Hz), 7.30 (2H, d, *J* = 7.6 Hz), 6.76 (1H, s), 6.68 (1H, d, *J* = 16.0 Hz), 3.86 (3H, s), 2.43 (3H, s); ¹³C NMR (CDCl₃) δ 166.6, 166.3, 163.3, 141.0, 130.1, 128.1, 127.1, 125.9, 123.7, 104.5, 52.5, 21.8; MS *m/z* 158(100), 243(M⁺); IR ν_{max} (cm⁻¹) 1711.8, 1651.1, 1560.4, 1528.8, 1429.6, 1385.3, 1312.4, 1262.2, 1173.4, 972.8, 816.9. Elemental analysis Calcd. for C₁₄H₁₃NO₃, C 69.12 %; H 5.39 %; N 5.76% Found C 68.98

%; H 5.52 %; N 5.89%.

5f: 100.3mg, pale yellow solid, mp. 76-78°C; ^1H NMR (CDCl_3) δ 7.69 (2H, d, $J = 6.8$ Hz), 7.30 (2H, d, $J = 6.8$ Hz), 6.65-6.59 (1H, m), 6.38 (1H, d, $J = 16.0$ Hz), 6.36 (1H, s), 1.97 (1H, d, $J = 7.2$ Hz); ^{13}C NMR (CDCl_3) δ 169.8, 162.0, 134.5, 132.5, 128.7, 128.6, 124.5, 117.4, 97.9, 19.1; MS m/z 69(100), 264(97, M^+), 266(94, $\text{M}^+ + 2$); IR ν_{max} (cm^{-1}) 1666.5, 1593.9, 155.9, 1501.8, 1424.2, 1376.0, 961.8, 846.8, 813.8, 774.1, 505.2. Elemental analysis Calcd. for $\text{C}_{12}\text{H}_{10}\text{BrNO}$, C 54.57 %; H 3.82 %; N 5.30% Found C 54.70 %; H 3.70 %; N 5.42%.

8c: 147.1mg, yellow solid, mp. 177-179°C; ^1H NMR (CDCl_3) δ 7.68 (2H, d, $J = 8.4$ Hz), 7.59 (4H, dd, $J_1 = 6.0$ Hz, $J_2 = 8.4$ Hz), 7.23 (2H, d, $J = 8.4$ Hz), 6.73 (1H, d, $J = 16.0$ Hz), 7.02 (1H, dd, $J_1 = 6.0$ Hz, $J_2 = 16.0$ Hz), 6.50 (1H, s), 5.39-5.36 (1H, m), 3.63 (1H, dd, $J_1 = 10.8$ Hz, $J_2 = 16.4$ Hz), 3.25 (1H, dd, $J_1 = 7.2$ Hz, $J_2 = 16.4$ Hz), 2.40 (3H, s); ^{13}C NMR (CDCl_3) δ 168.3, 162.2, 156.7, 141.1, 134.8, 132.6, 130.0, 128.7, 128.2, 127.1, 126.7, 124.8, 117.3, 100.5, 80.3, 41.5, 21.9; MS m/z 131(100), 408($\text{M}^+ - 1$), 410($\text{M}^+ + 1$); IR ν_{max} (cm^{-1}) 2916.7, 1608.2, 1596.1, 1561.4, 1430.4, 1366.2, 967.9, 899.1, 819.0, 803.8. Elemental analysis Calcd. for $\text{C}_{21}\text{H}_{17}\text{BrN}_2\text{O}_2$, C 61.63 %; H 4.19 %; N 6.84% Found C 61.77 %; H 4.07 %; N 6.72%.

8d: 118.2mg, yellow solid, mp. 133-134°C; ^1H NMR (CDCl_3) δ 7.78 (2H, m), 7.57 (2H, d, $J = 8.0$ Hz), 7.22 (2H, d, $J = 8.0$ Hz), 7.14 (2H, t, $J = 8.4$ Hz), 6.72 (1H, d, $J = 16.0$ Hz), 6.61 (1H, dd, $J_1 = 6.0$ Hz, $J_2 = 16.0$ Hz), 6.47 (1H, s), 5.39-5.33 (1H, m), 3.63 (1H, dd, $J_1 = 11.2$ Hz, $J_2 = 16.4$ Hz), 3.24 (1H, dd, $J_1 = 8.0$ Hz, $J_2 = 16.4$ Hz), 2.38 (3H, s); ^{13}C NMR (CDCl_3) δ 167.7, 163.7($J = 248.6$ Hz), 161.8, 156.3, 140.7, 134.2, 129.5, 128.7($J = 8.1$ Hz), 126.7, 126.3, 125.1($J = 2.4$ Hz), 117.0, 116.0($J = 21.4$ Hz), 110.1, 79.9, 41.0, 21.4; MS m/z 162(100), 348(M^+); IR ν_{max} (cm^{-1}) 2962.1, 1608.2, 1560.3, 1430.2, 1366.3, 966.9, 898.6, 831.2, 797.6. Elemental analysis Calcd. for $\text{C}_{21}\text{H}_{17}\text{FN}_2\text{O}_2$, C 72.40 %; H 4.92 %; N 8.04% Found C 72.20 %; H 4.80 %; N 8.14%.

8j: 108.5mg, pale yellow solid, mp. 138-139°C; ^1H NMR (CDCl_3) δ 7.78 (2H, m), 7.54

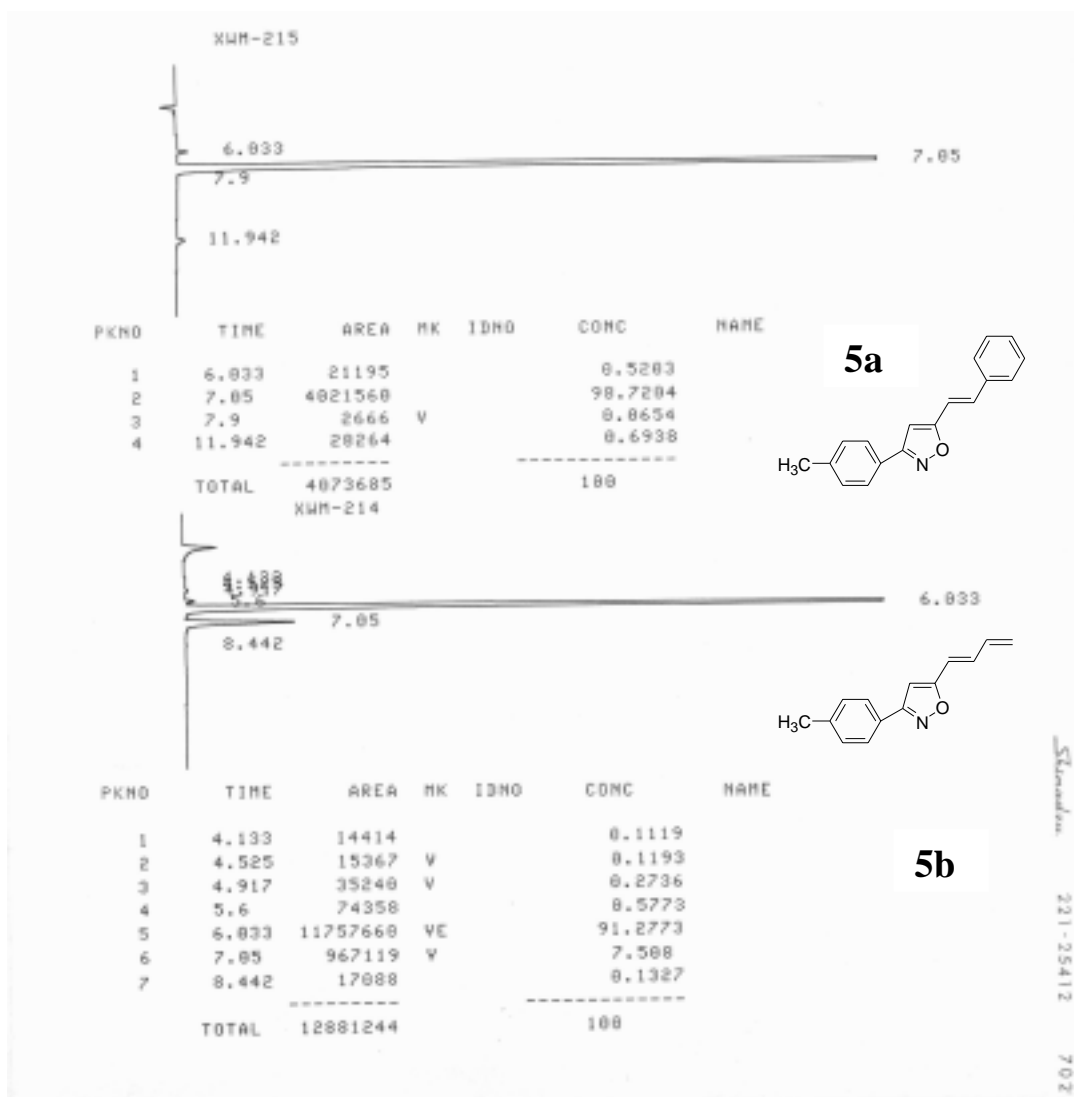
(2H, d, $J = 8.2$ Hz), 7.20 (2H, d, $J = 8.2$ Hz), 7.13 (2H, t, $J = 8.4$ Hz), 6.70 (1H, d, $J = 16.0$ Hz), 6.67 (1H, d, $J = 16.0$ Hz), 6.43 (1H, s), 3.36 (1H, d, $J = 16.4$ Hz), 3.27 (1H, d, $J = 16.4$ Hz), 2.37 (3H, s), 1.68 (3H, s); ^{13}C NMR (CDCl_3) δ 168.0, 163.7($J = 249.1$ Hz), 161.8, 156.2, 140.5, 138.6, 129.4, 128.7($J = 8.2$ Hz), 126.7, 126.5, 125.2($J = 3.6$ Hz), 116.0($J = 22.0$ Hz), 114.2, 100.0, 85.5, 46.7, 25.4, 21.4; MS m/z 162(100), 362(M^+); IR ν_{max} (cm^{-1}) 2924.2, 1607.4, 1560.7, 1429.6, 1378.4, 1365.1, 964.7, 896.8, 832.0, 796.0; Elemental analysis Calcd. for $\text{C}_{22}\text{H}_{19}\text{FN}_2\text{O}_2$, C 72.91 %; H 5.28 %; N 7.73 % Found C 72.70 %; H 5.39 %; N 7.82%.

8k: 137.9mg, pale yellow solid, mp. 115-116°C; ^1H NMR (CDCl_3) δ 7.79-7.77 (2H, m), 7.59-7.54 (4H, m), 7.46-7.40 (6H, m), 7.21 (2H, d, $J = 8.4$ Hz), 6.92 (1H, d, $J = 16.0$ Hz), 6.69 (1H, d, $J = 16.0$ Hz), 6.48 (1H, s), 3.76 (2H, s), 2.38 (3H, s); ^{13}C NMR (CDCl_3) δ 167.7, 162.7, 156.2, 141.7, 140.7, 137.8, 130.0, 129.5, 129.4, 128.9, 128.1, 126.7, 126.6, 126.4, 125.6, 115.2, 100.5, 89.2, 47.8, 21.4; MS m/z 143(100), 406(M^+); IR ν_{max} (cm^{-1}) 3024.1, 2924.2, 1599.1, 1560.3, 1430.7, 1366.6, 968.8, 900.4, 831.0, 801.8. Elemental analysis Calcd. for $\text{C}_{27}\text{H}_{22}\text{N}_2\text{O}_2$, C 79.78 %; H 5.46 %; N 6.89 % Found C 79.97 %; H 5.31 %; N 6.76 %.

HPLC copys of **5a-b**, **8b-e**, **8g-h** (page 6-9).

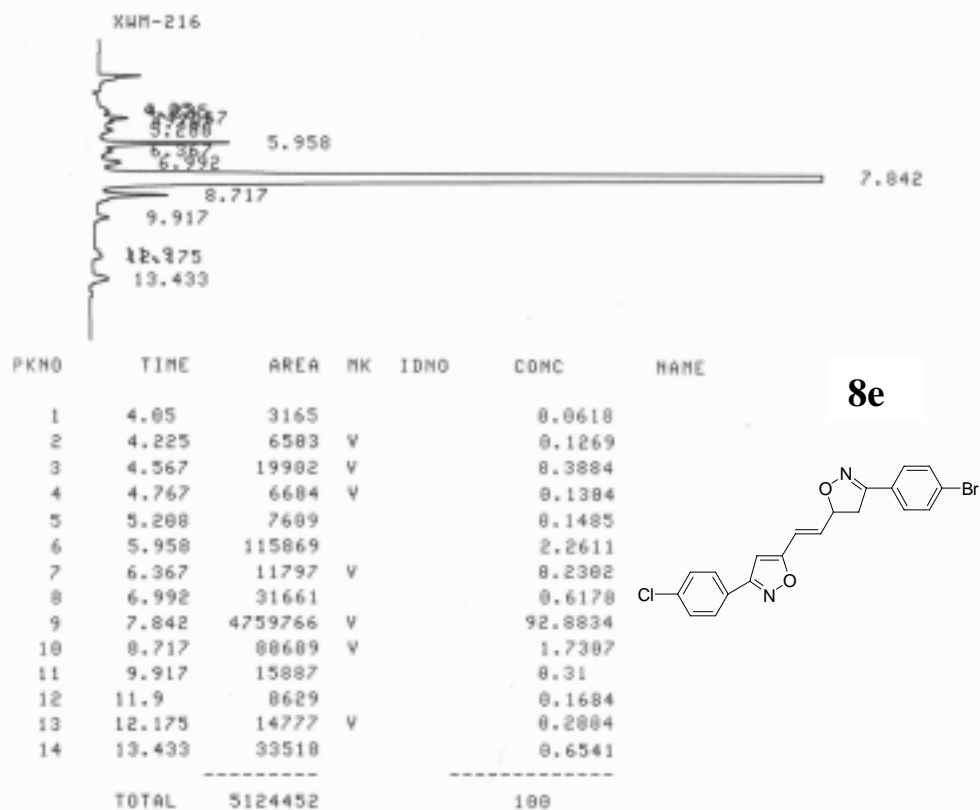
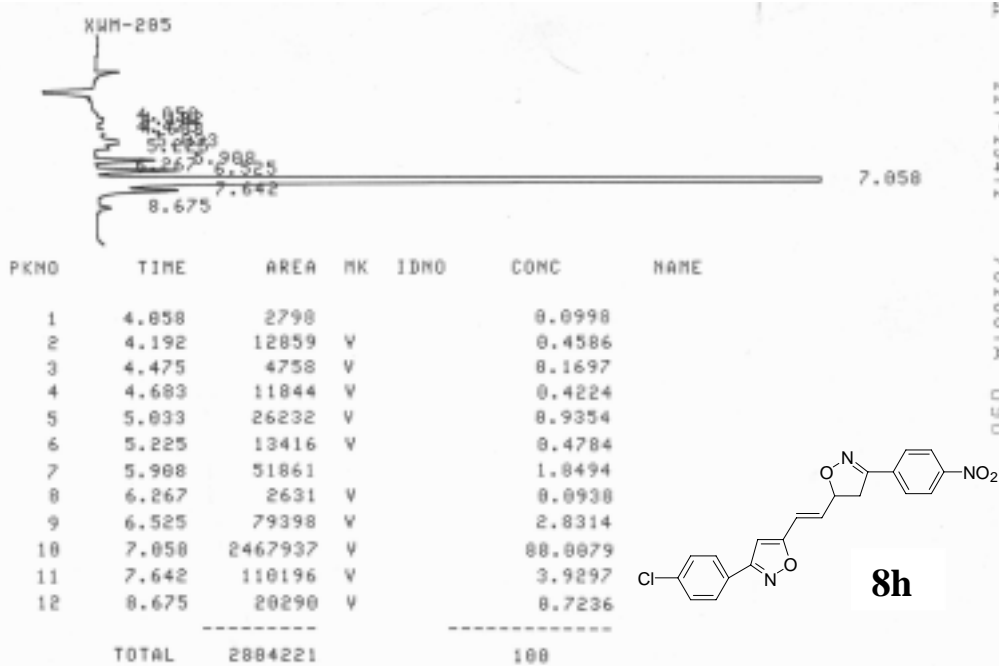
^1H , ^{13}C NMR copys of **5a-c**, **8a**, **8c-f**, **8j-k** (page 10-29).

HPLC copys of **5a** and **5b**

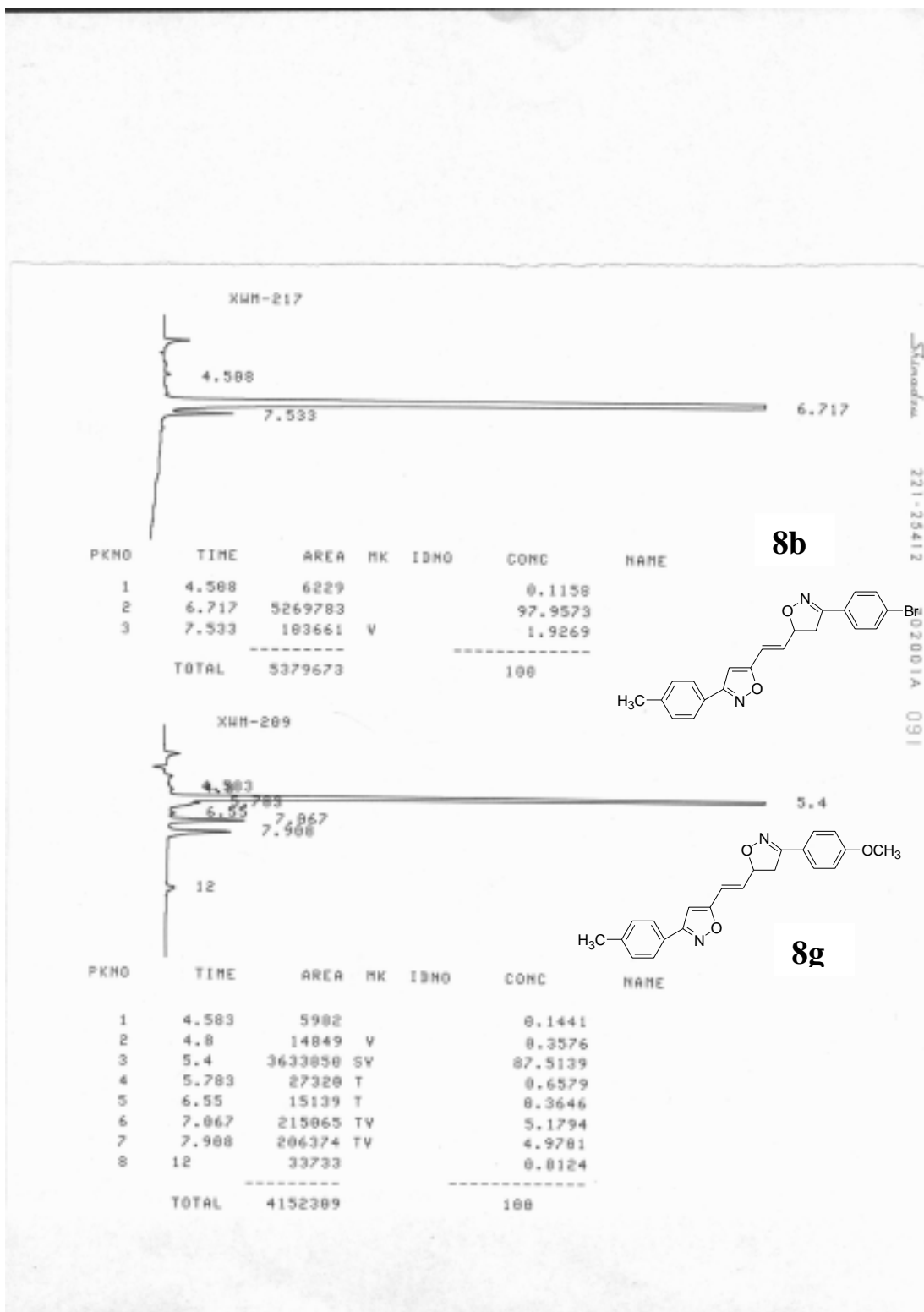


HPLC copys of 8h and 8e

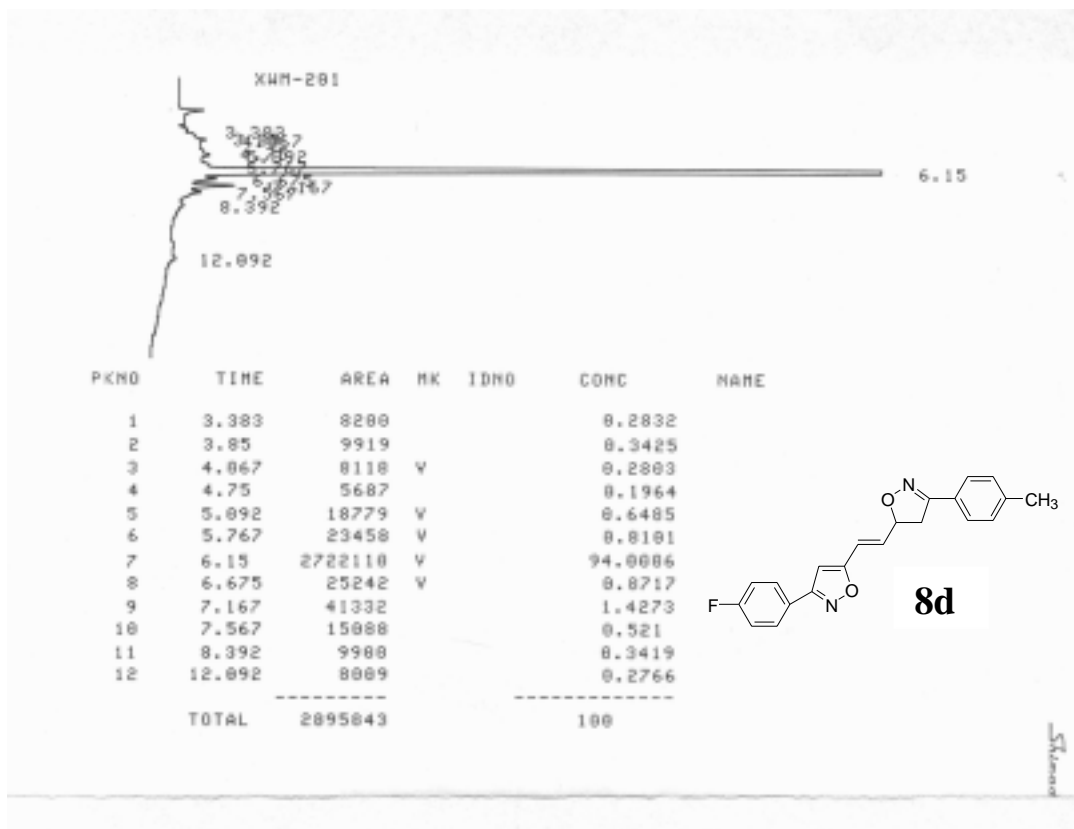
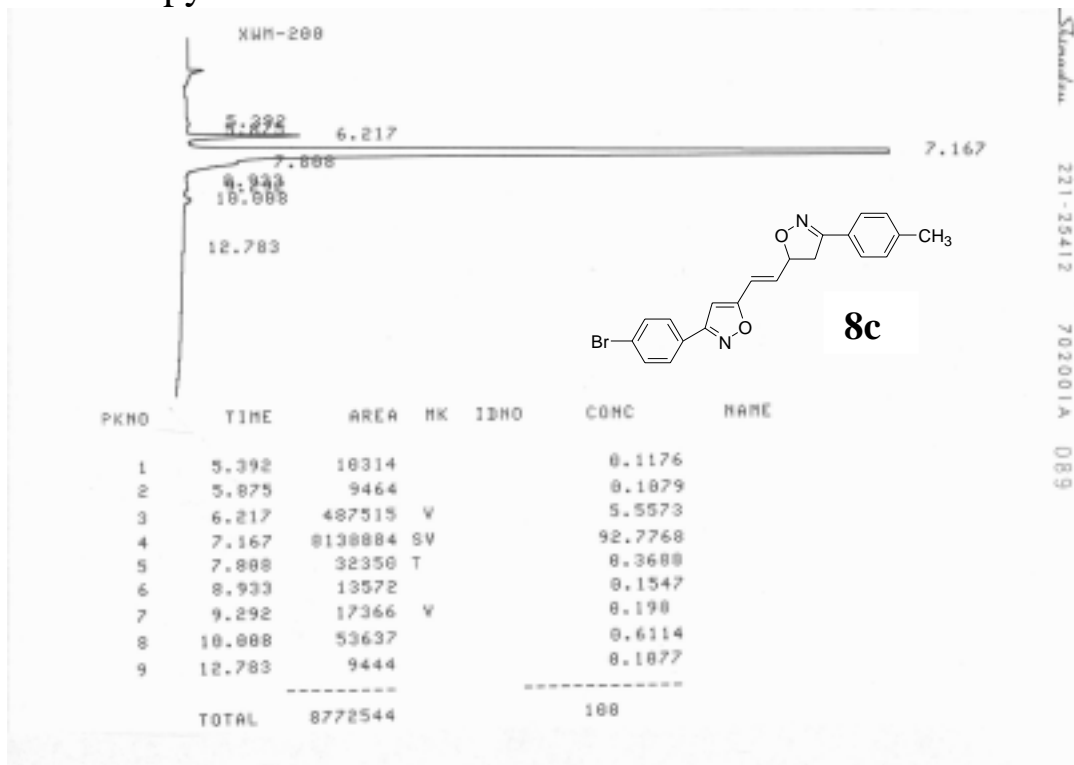
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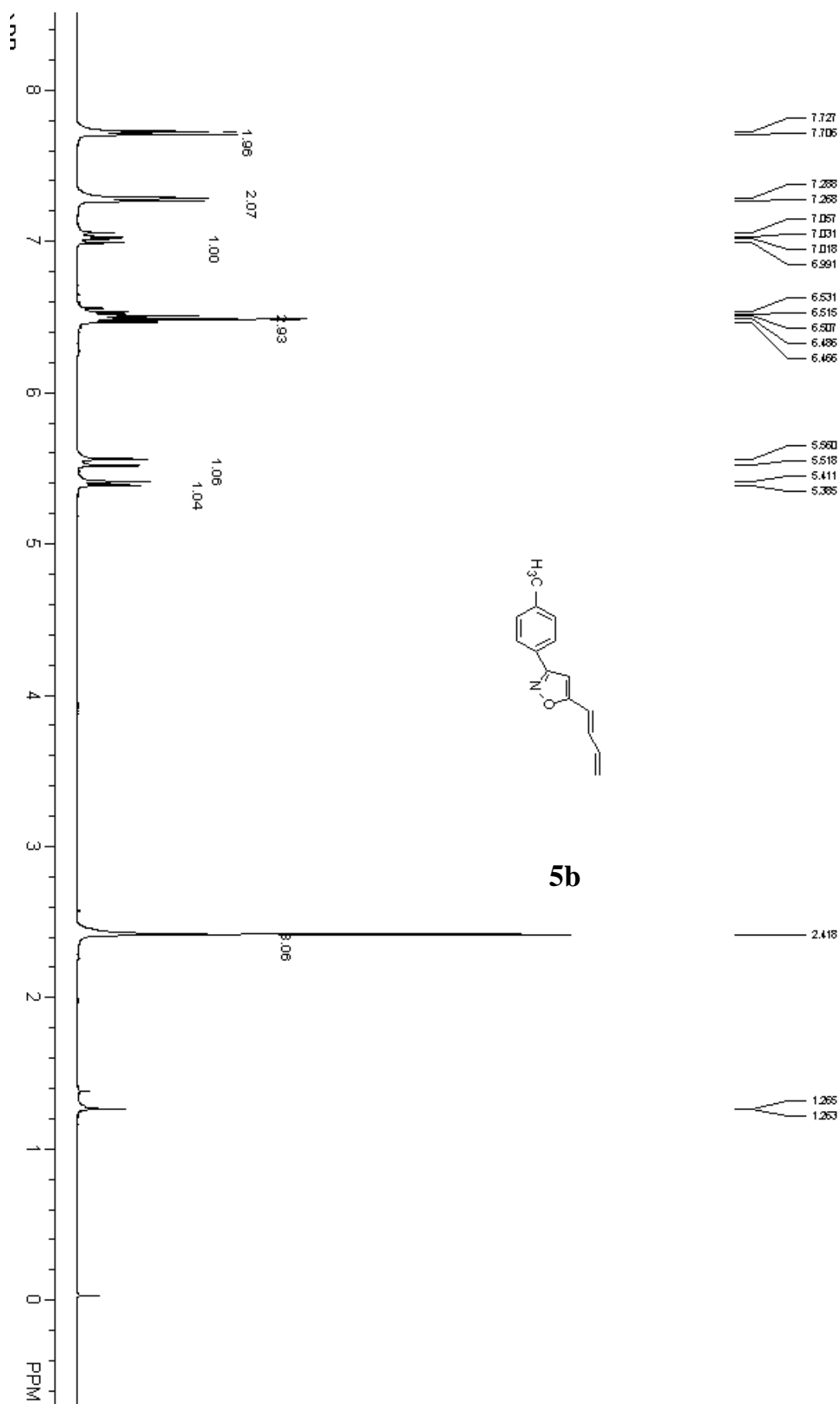


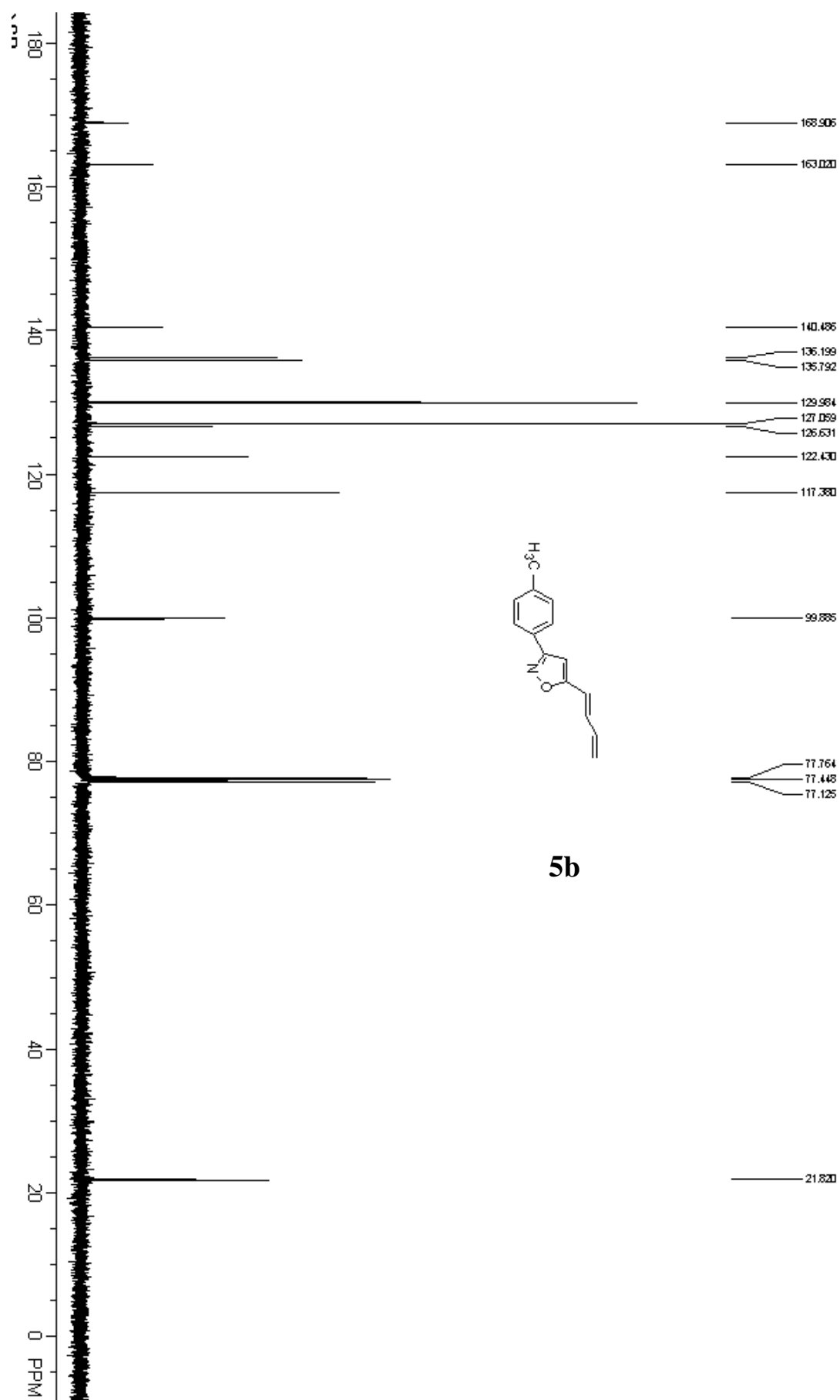
HPLC copys of 8b and 8g



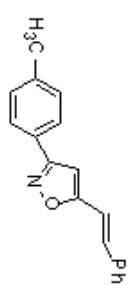
HPLC copys of 8c and 8d







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7.380
7.369
7.341
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7.262
7.016
6.975
6.544



5a

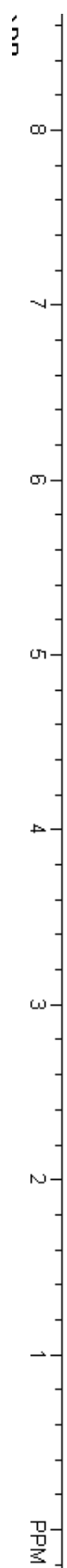
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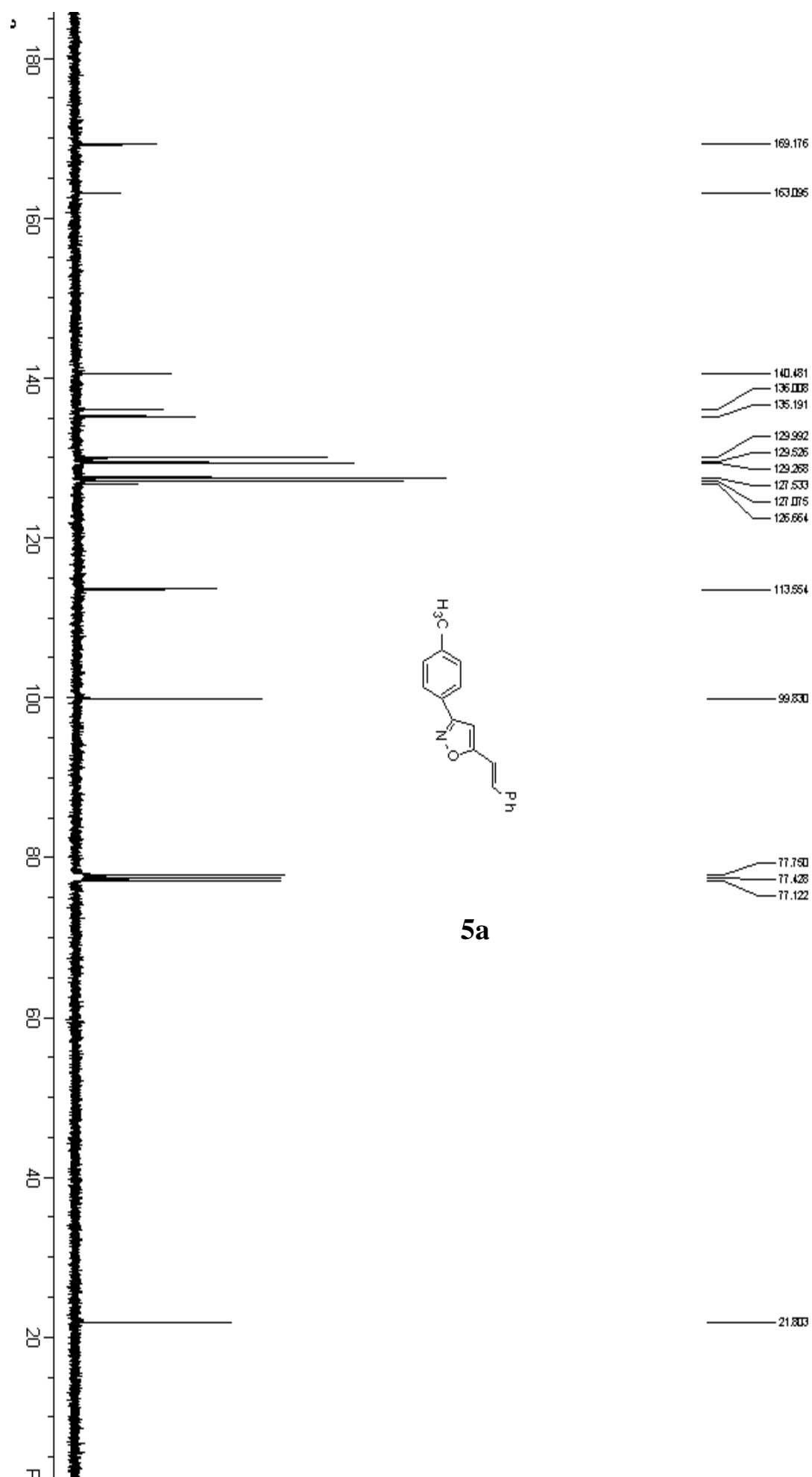
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4.19

2.08 2.00 2.32

1.05 1.00



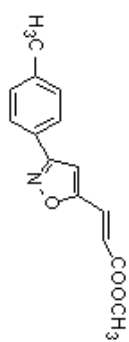


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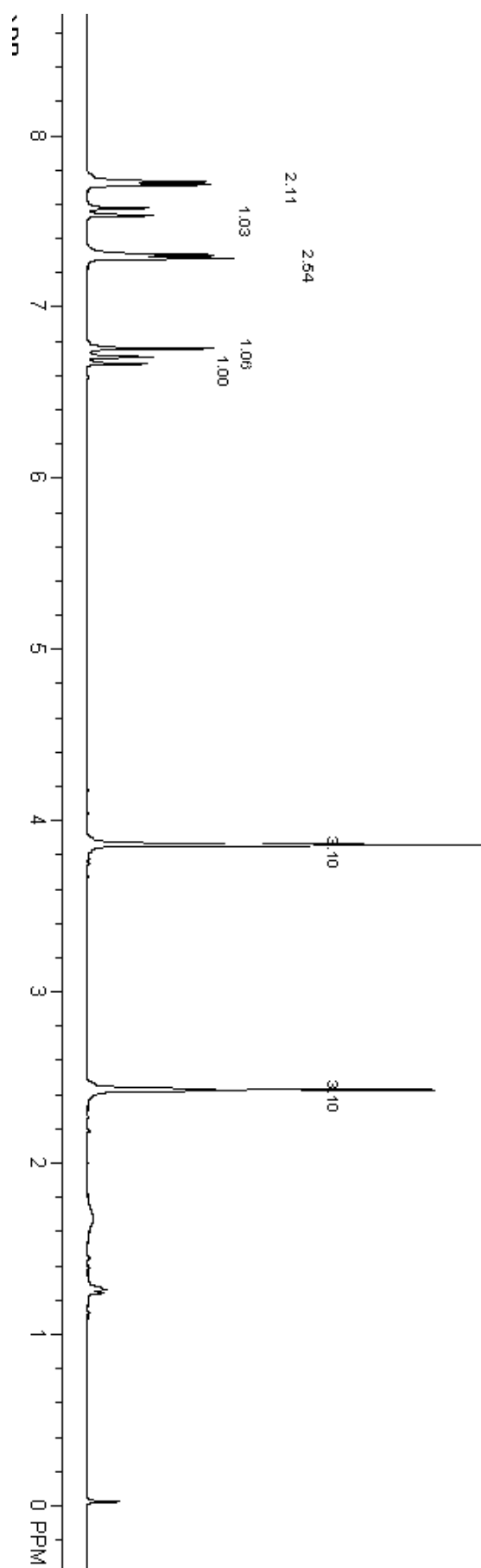
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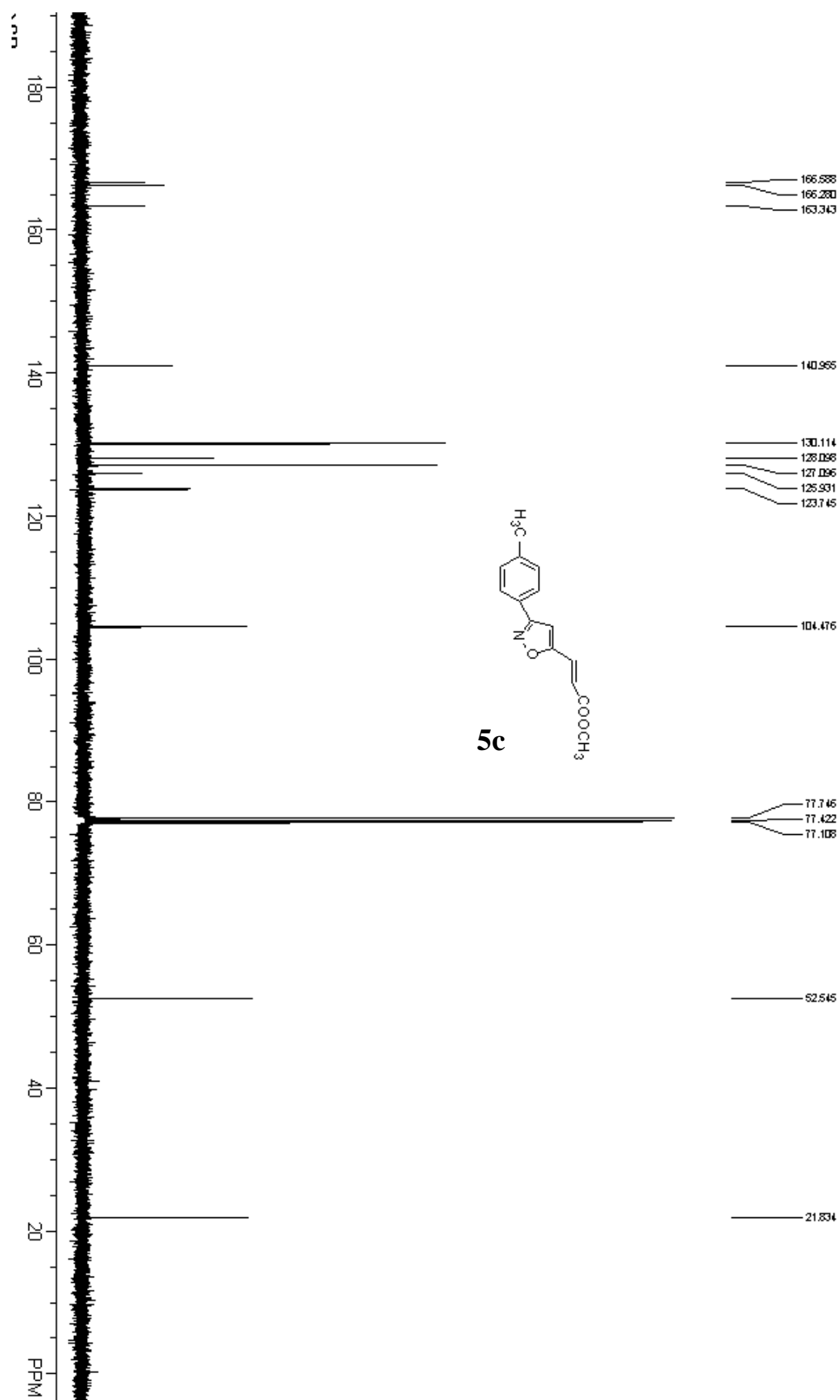
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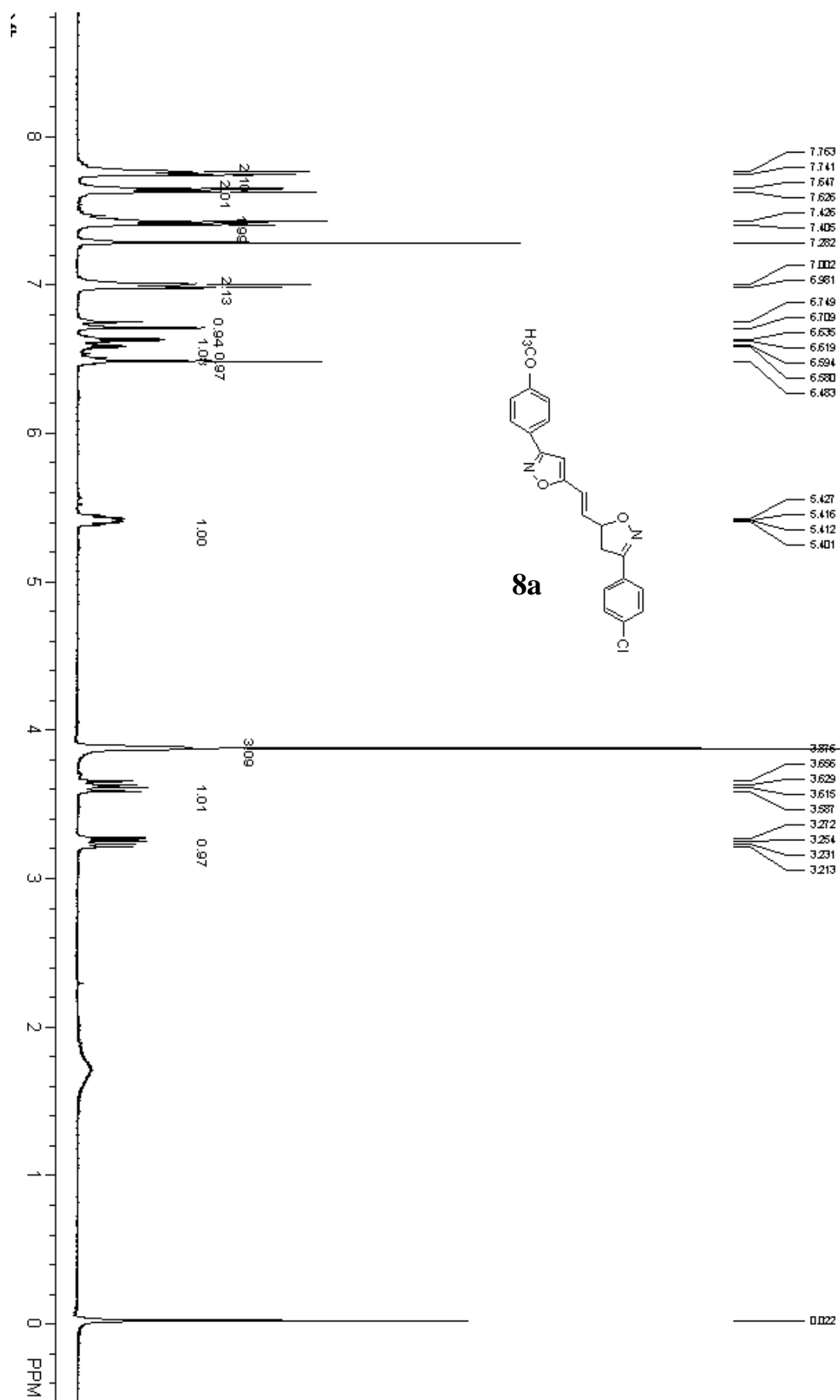
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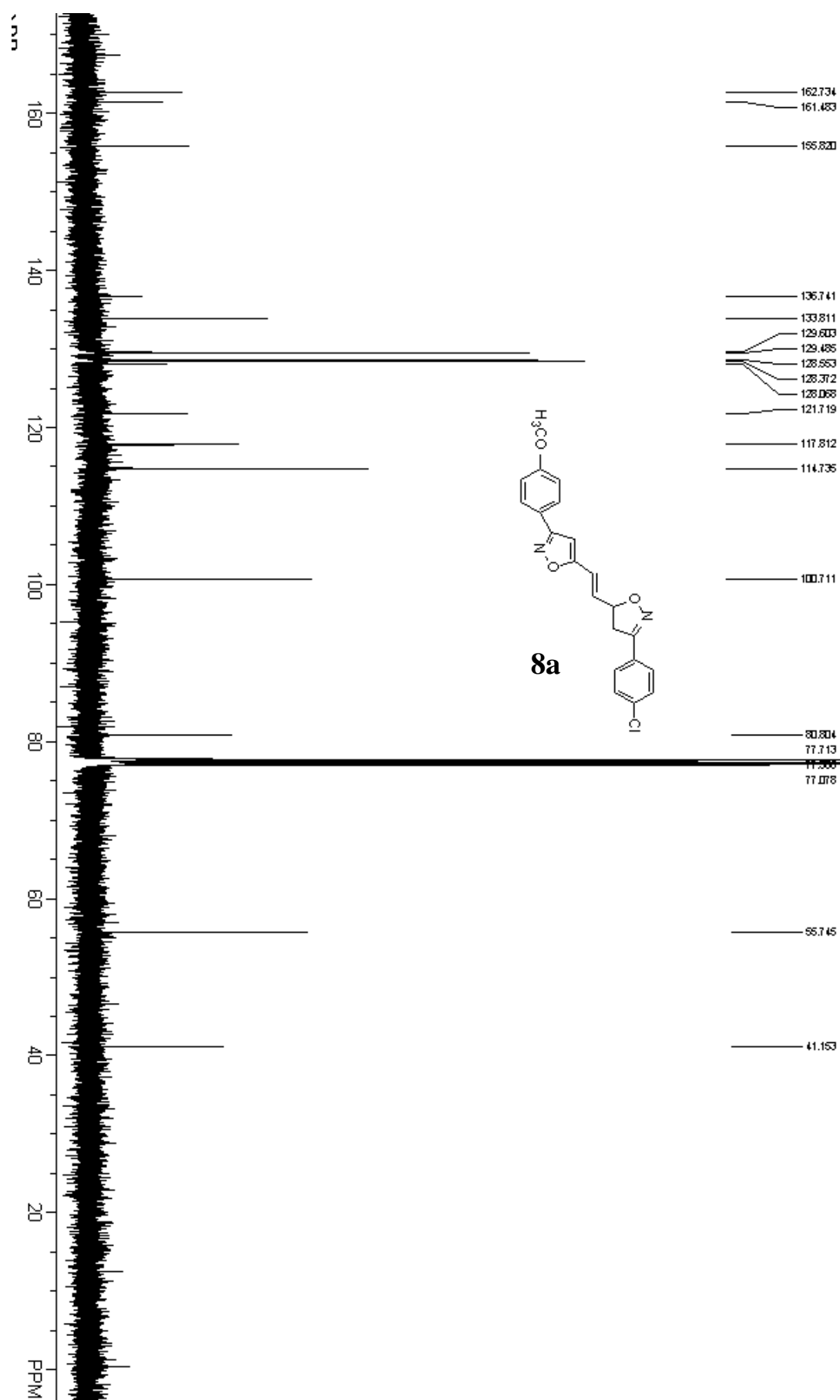


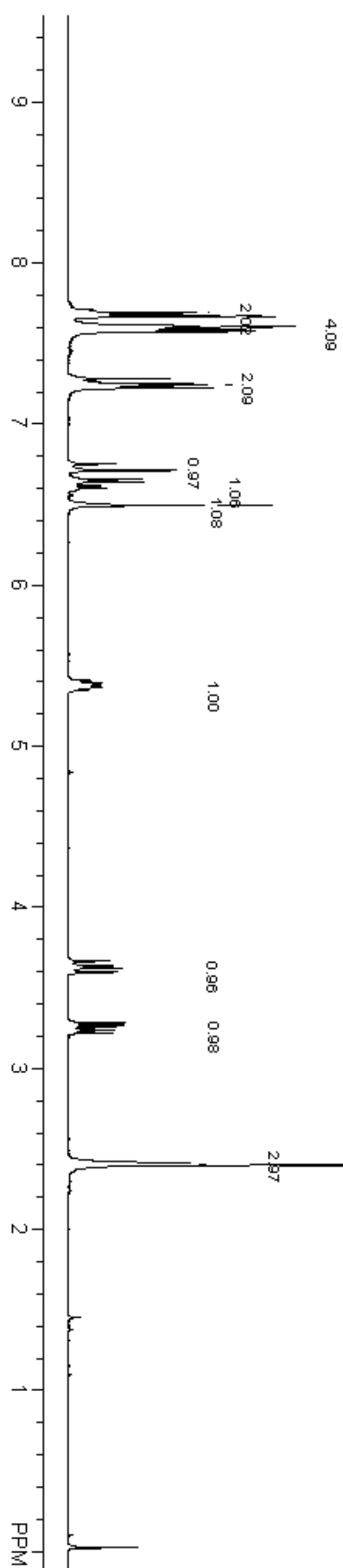
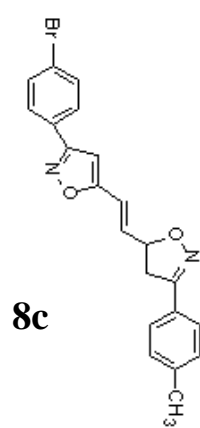
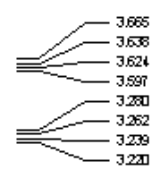
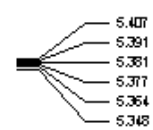
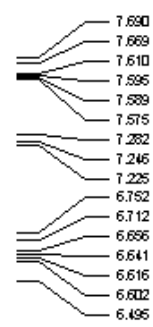
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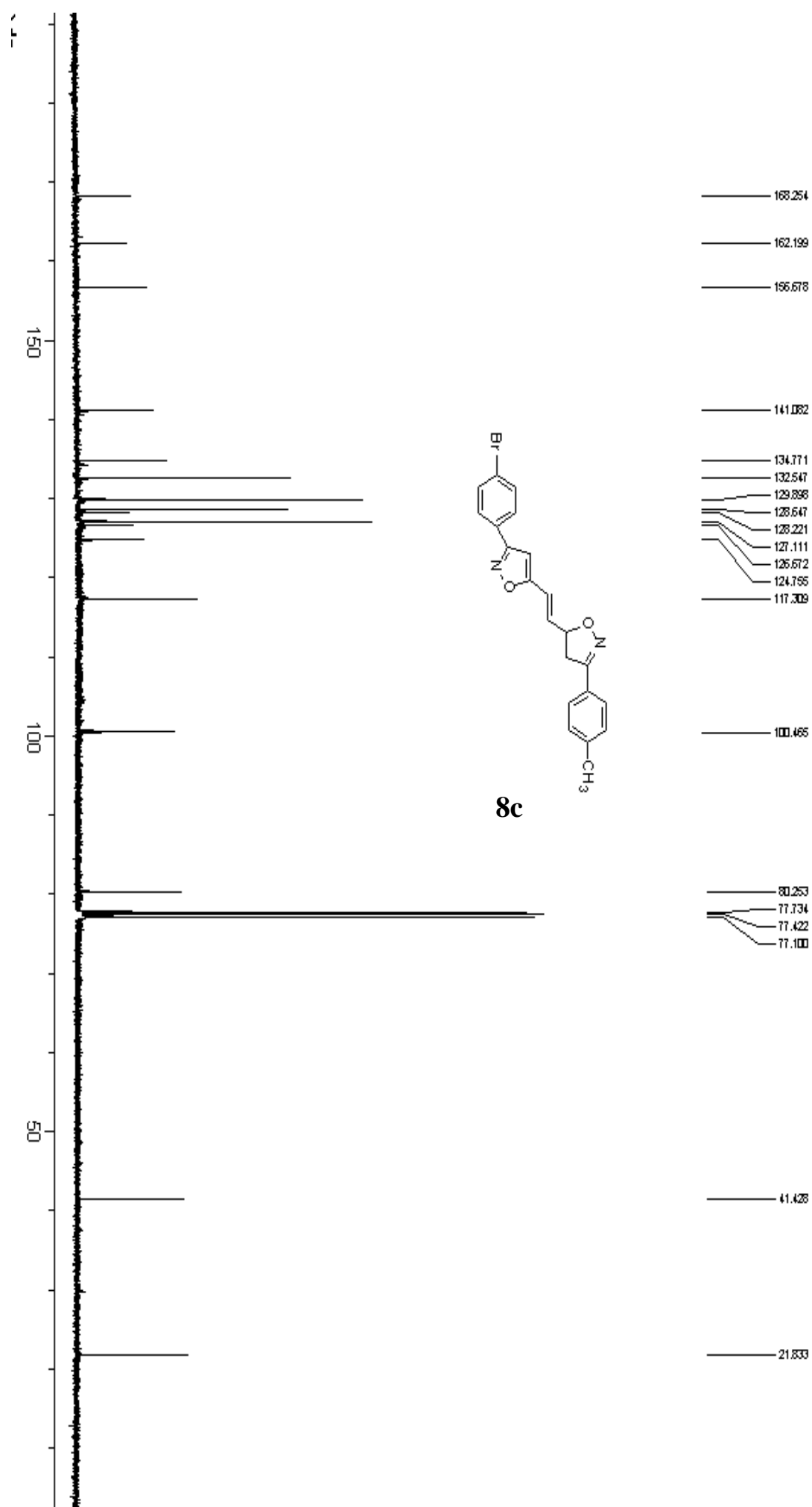


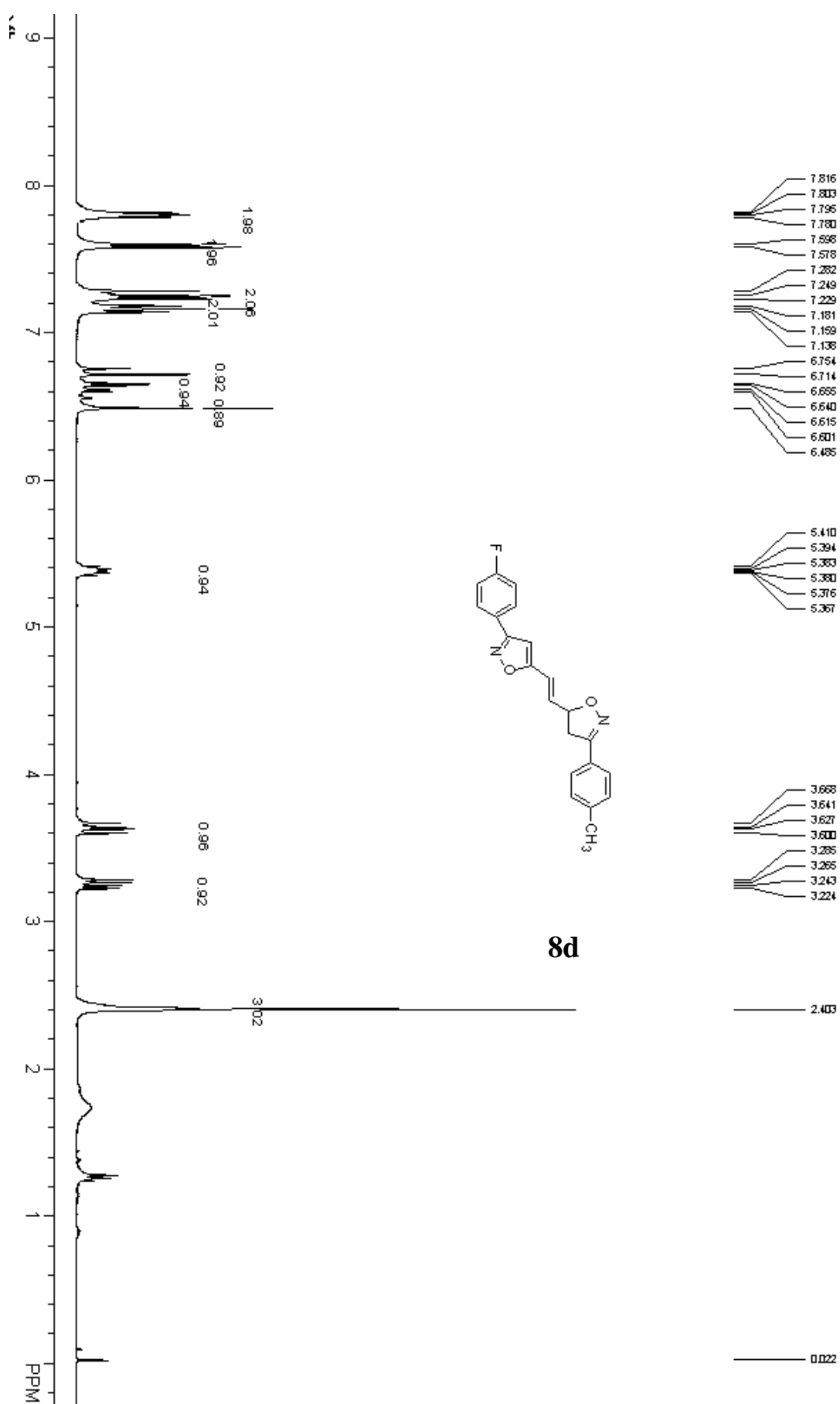


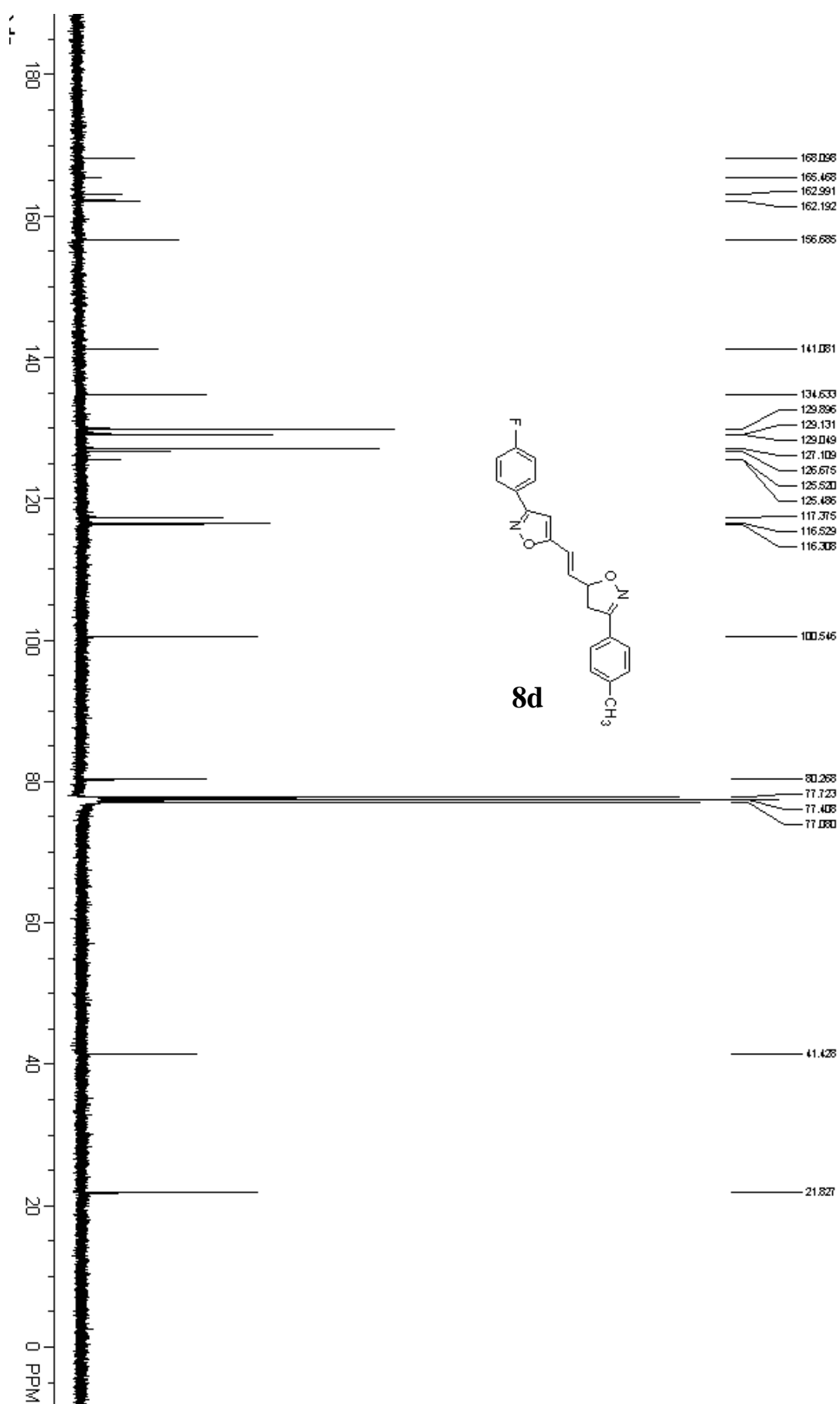


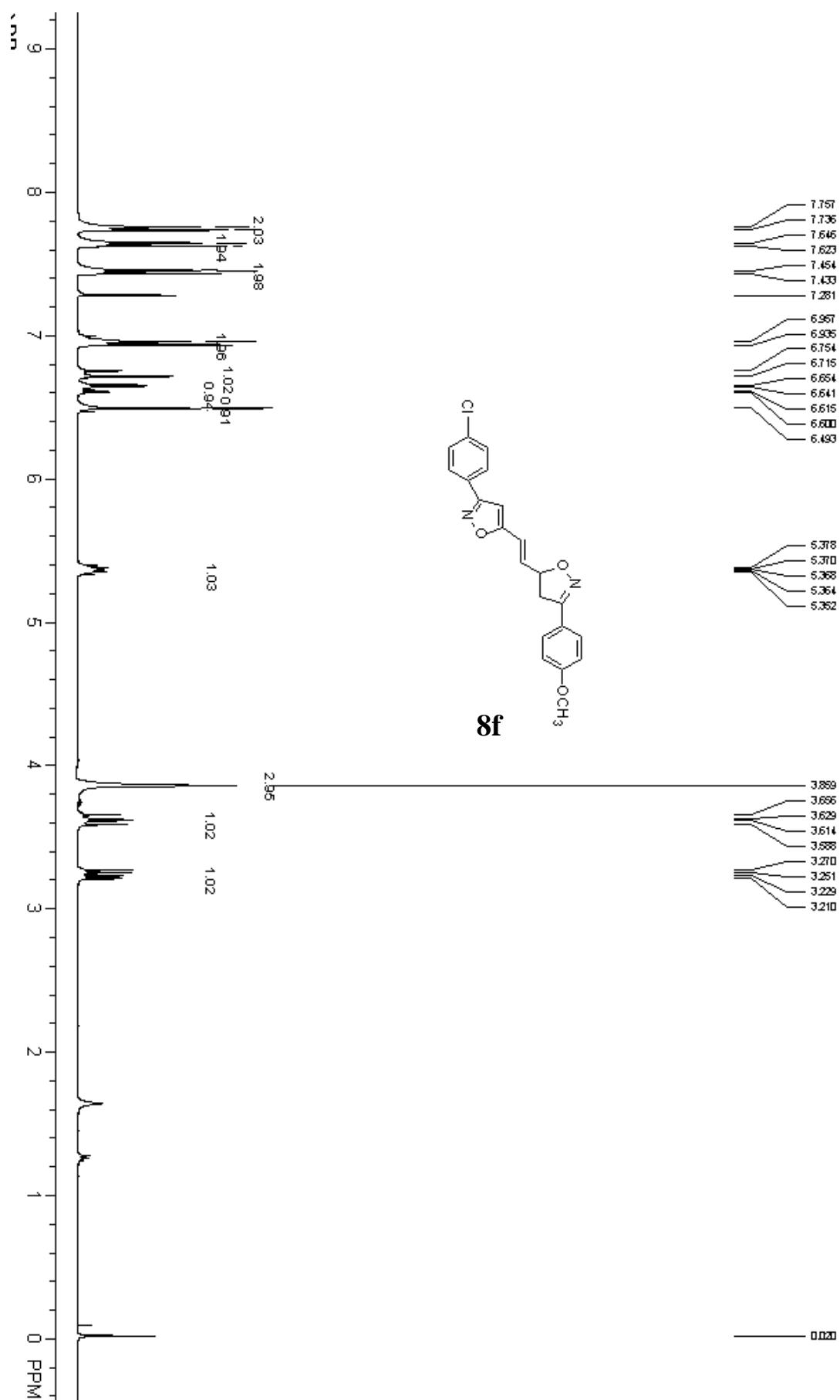


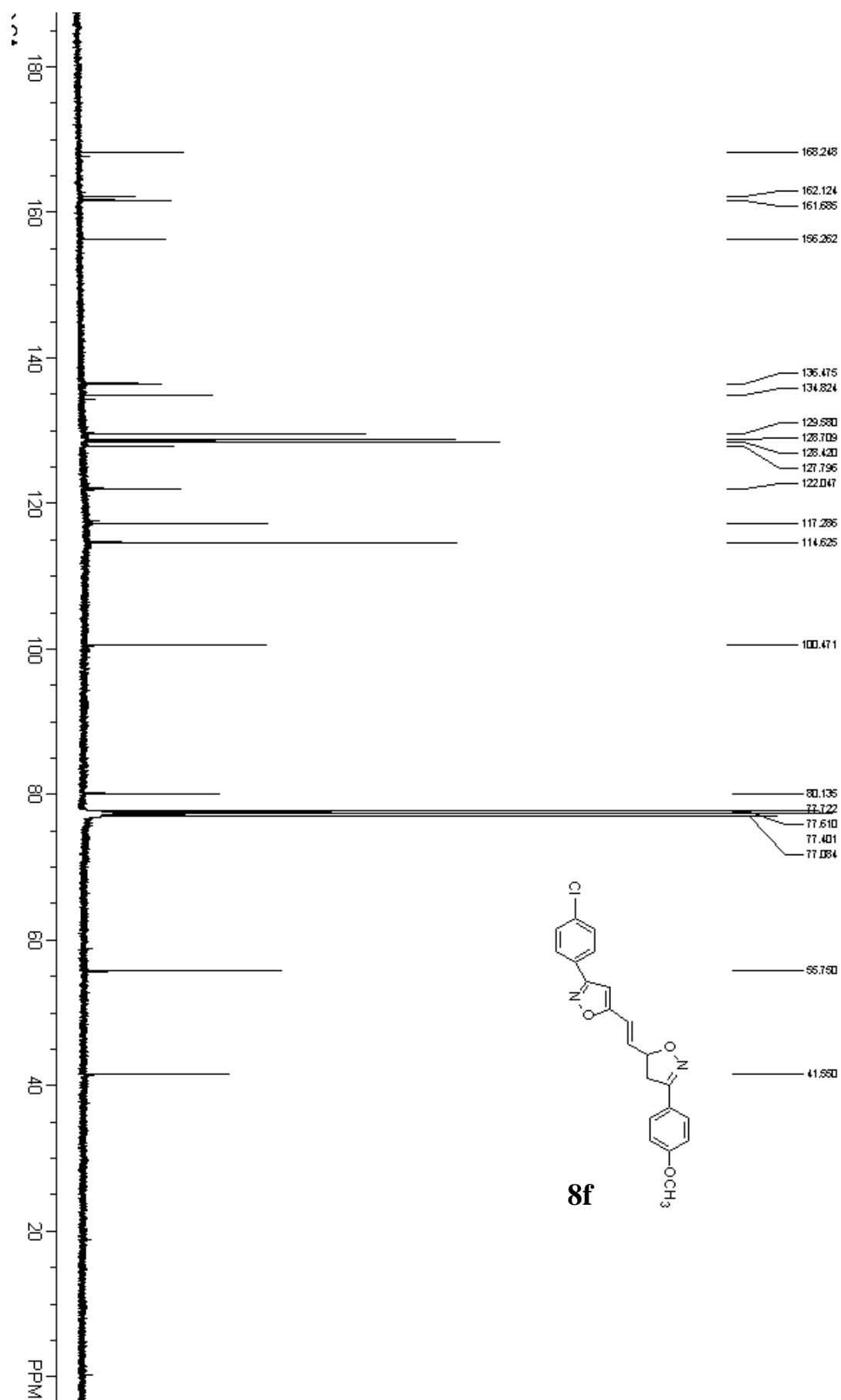


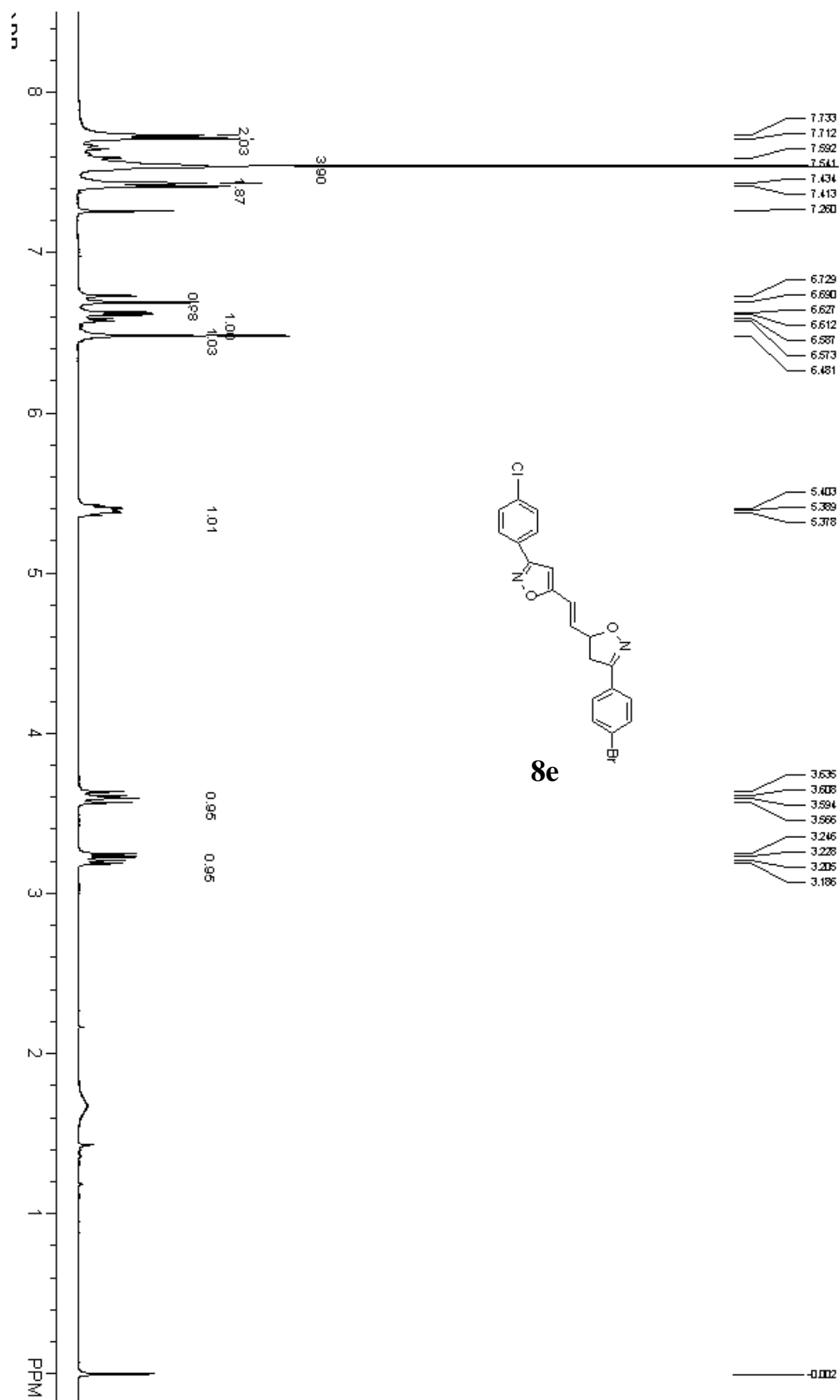


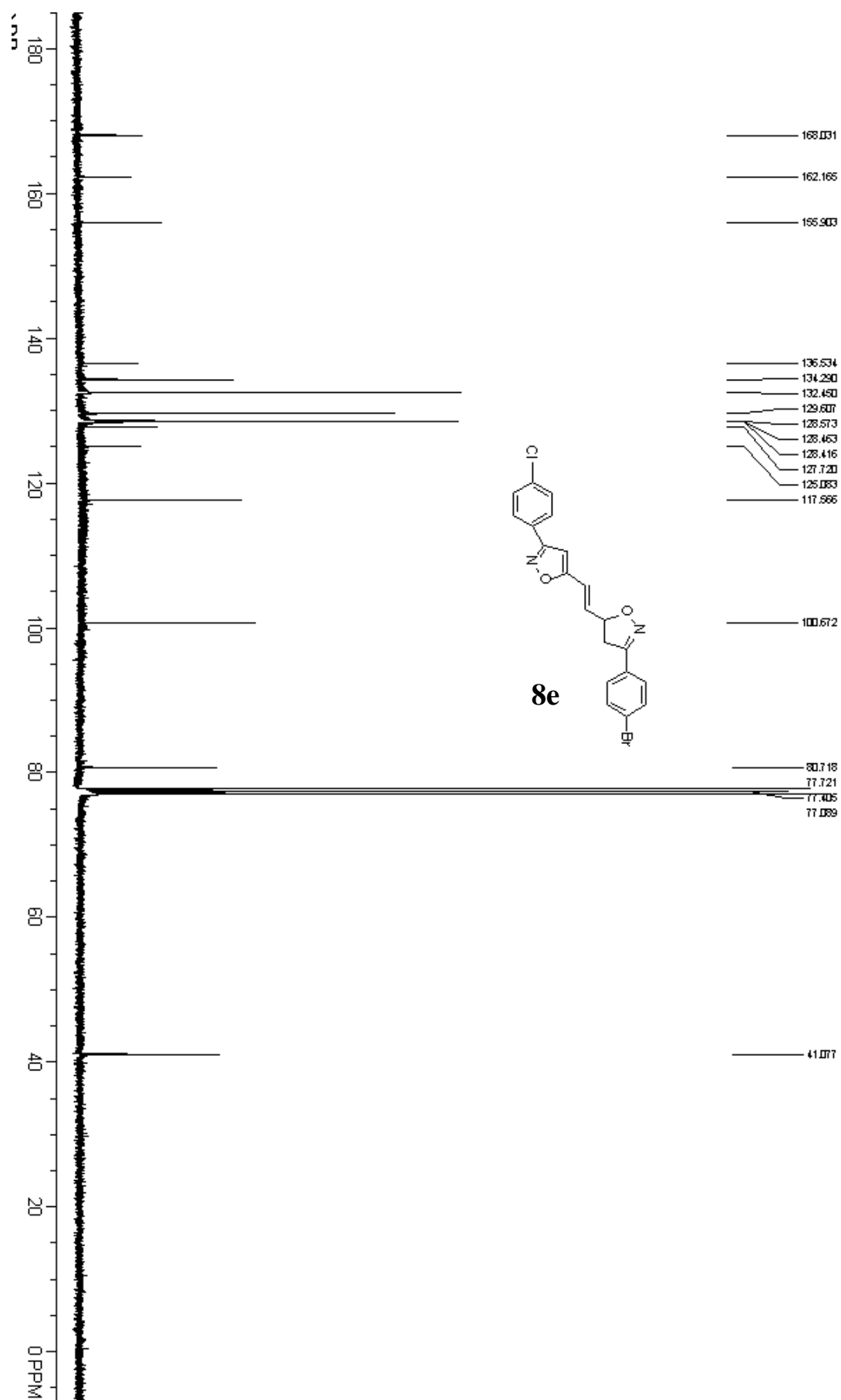


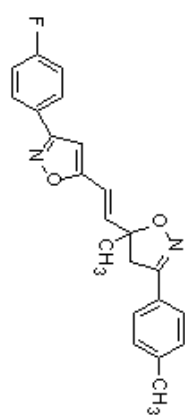
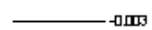
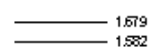
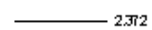
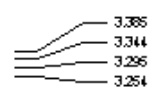
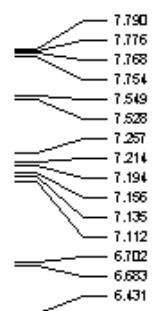




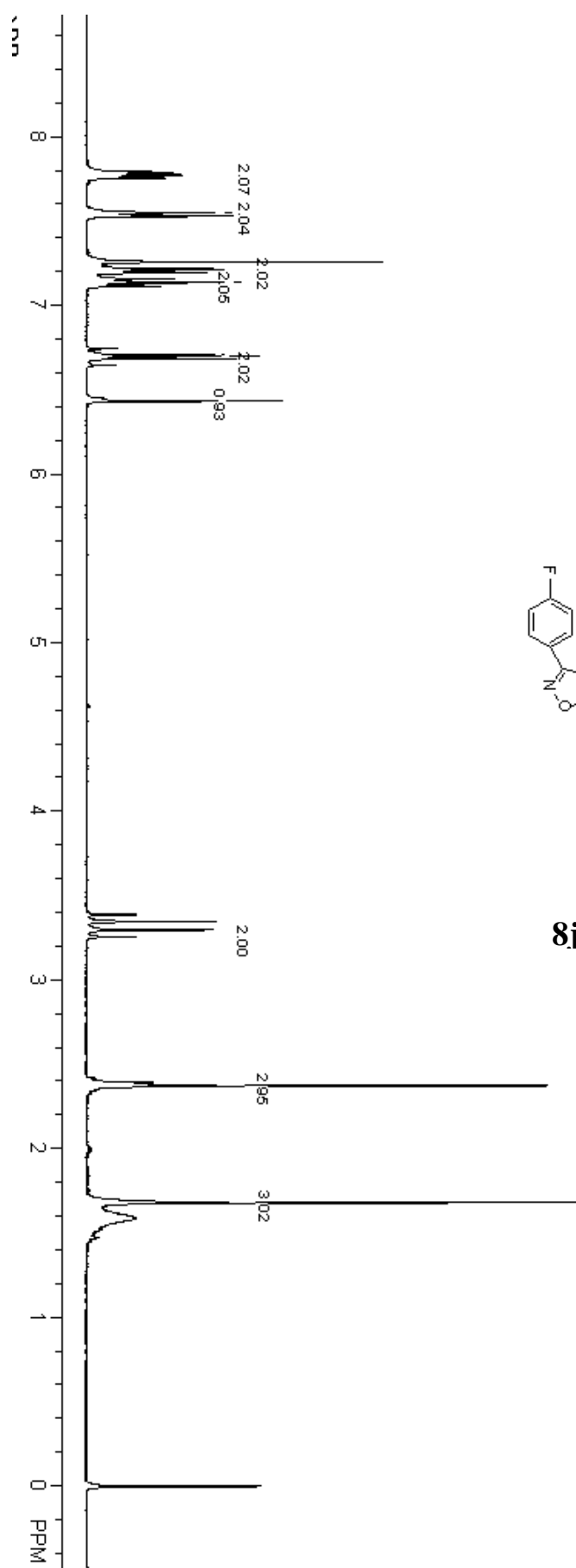


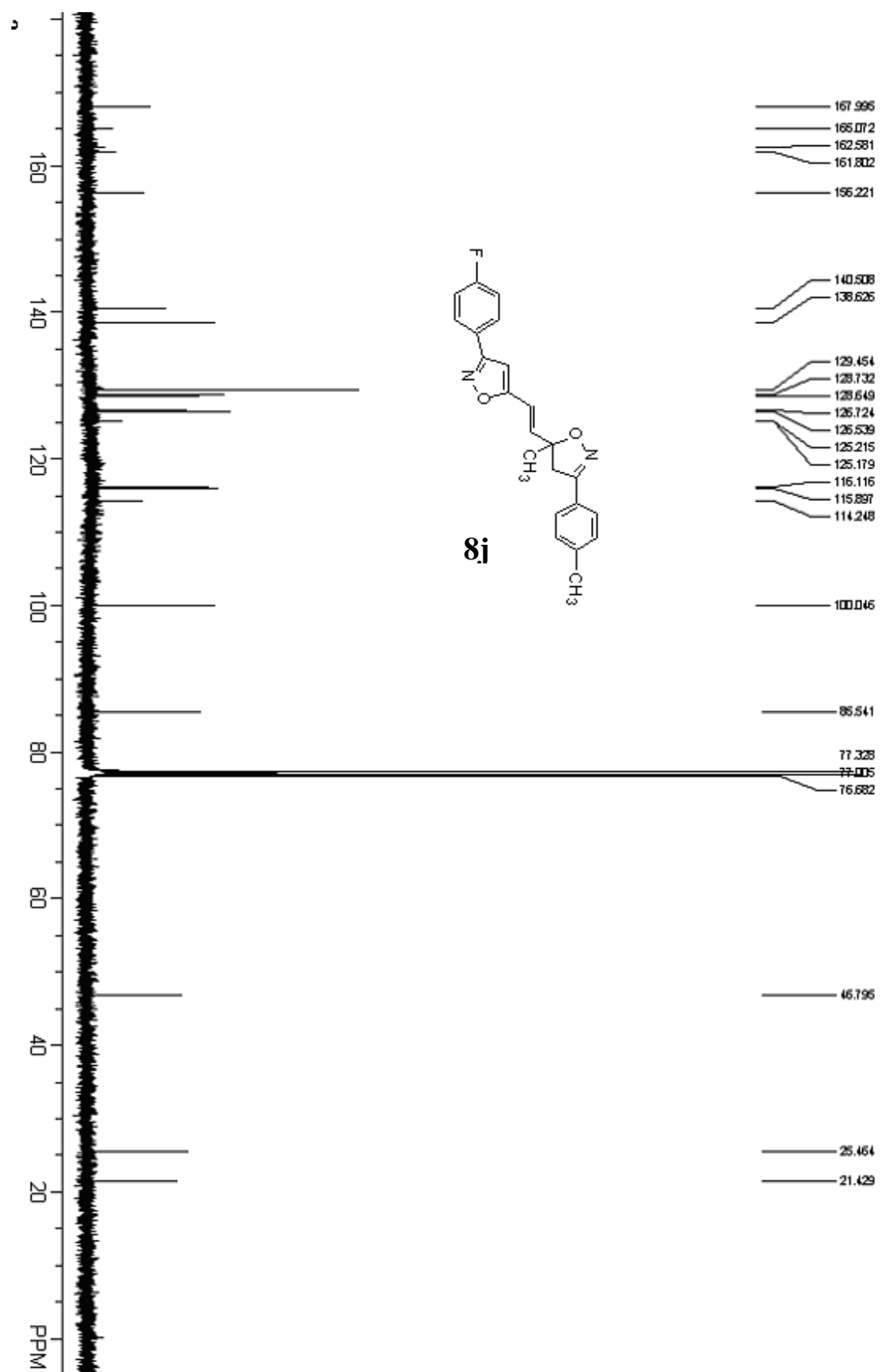






8j



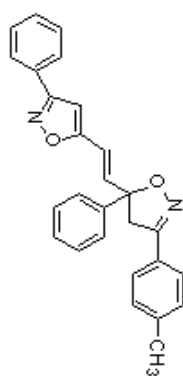


7.790
7.780
7.772
7.767
7.589
7.568
7.543
7.451
7.447
7.441
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6.895
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6.669
6.479

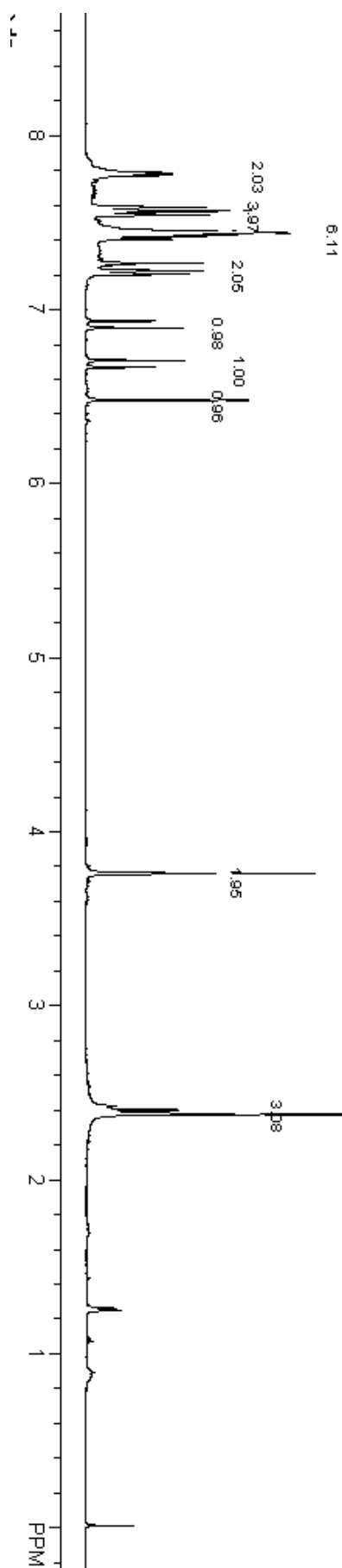
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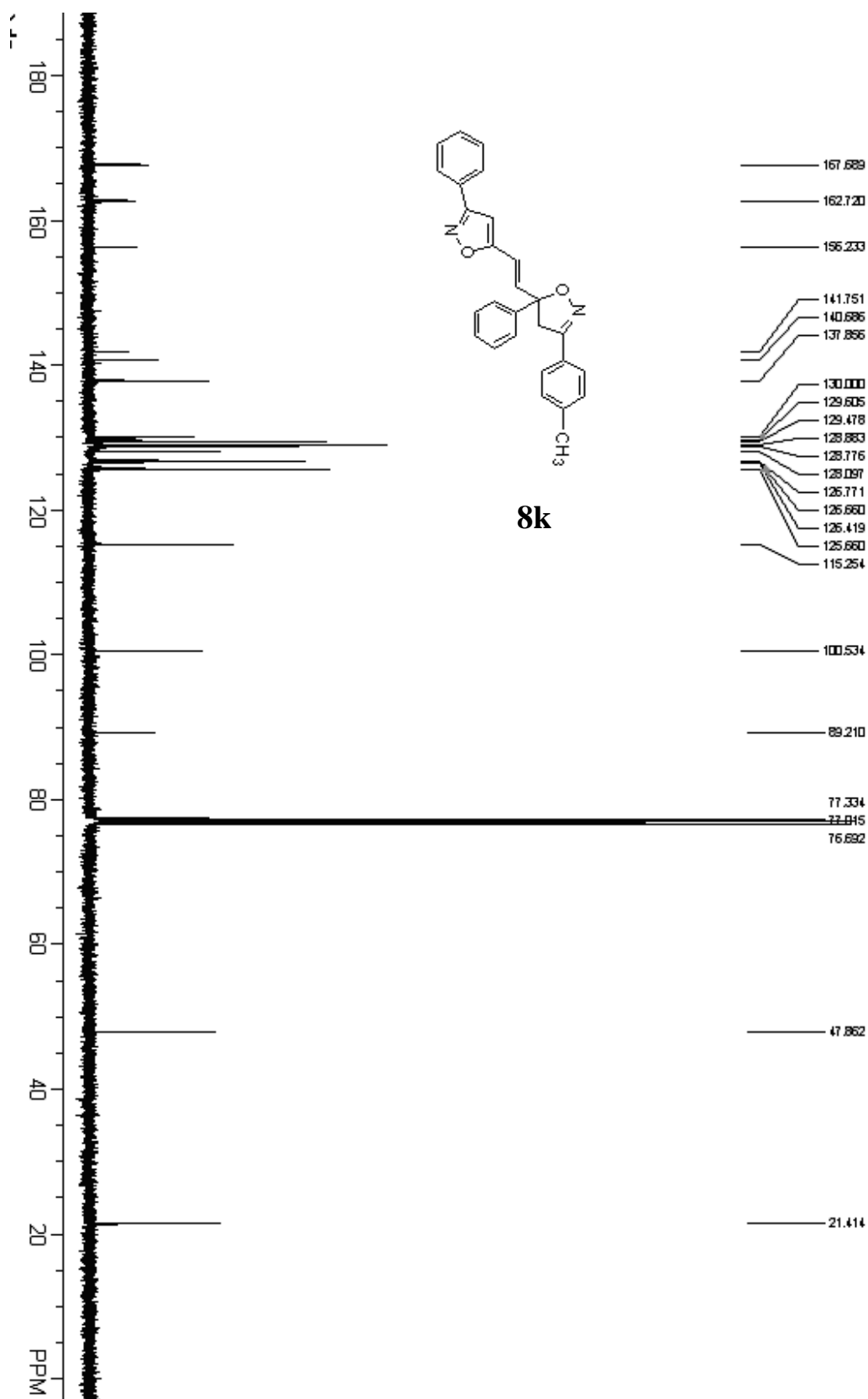
2.377

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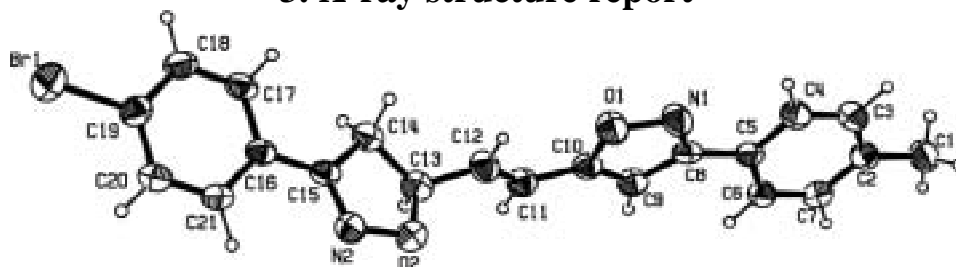


8k





3. X-ray structure report



Crystal Data

Empirical Formula	C ₂₁ H ₁₇ N ₂ BrO ₂
Formula Weight	409.28
Crystal Color, Habit	yellow, platelet
Crystal Dimensions	0.45 X 0.25 X 0.16 mm
Crystal System	monoclinic
Lattice Type	Primitive
Detector Position	127.40 mm
Pixel Size	0.100 mm
Space Group	P2 ₁ /c (#14)
Z value	4
D _{calc}	1.476 g/cm ³
F ₀₀₀	832.00
Diffractometer	Rigaku RAXIS-RAPID